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# Development of a Multiple-Locus Variable-Number Tandem-Repeat Typing Scheme for Genetic Fingerprinting of *Burkholderia cenocepacia* and Application to Nationwide Epidemiological Analysis

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Organisms of the *Burkholderia cepacia* complex are especially important pathogens in cystic fibrosis (CF), with a propensity for patient-to-patient spread and long-term respiratory colonization. *B. cenocepacia* and *Burkholderia multivorans* account for the majority of infections in CF, and major epidemic clones have been recognized throughout the world. The aim of the present study was to develop and evaluate a multilocus variable-number tandem-repeat (VNTR) analysis (MLVA) scheme for *B. cenocepacia*. Potential VNTR loci were identified upon analysis of the annotated genome sequences of *B. cenocepacia* strains AU1054, J2315, and MCO-3, and 10 of them were selected on the basis of polymorphisms and size. A collection of 100 *B. cenocepacia* strains, including epidemiologically related and unrelated strains, as well as representatives of the major epidemic lineages, was used to evaluate typeability, epidemiological concordance, and the discriminatory power of MLVA-10 compared with those of pulsed-field gel electrophoresis (PFGE) and multilocus sequence typing (MLST). Longitudinal stability was assessed by testing 39 successive isolates from 14 patients. Typeability ranged from 0.91 to 1, except for that of one marker, which was not amplified in 53% of the *B. cenocepacia* IIIA strains. The MLVA types were shown to be stable in chronically colonized patients and within outbreak-related strains, with excellent epidemiological concordance. Epidemic and/or globally distributed lineages (epidemic Edinburgh-Toronto electrophoretic type 12 [ET-12], sequence type 32 [ST-32], ST-122, ST-234, and ST-241) were successfully identified. Conversely, the discriminatory power of MLVA was lower than that of PFGE or MLST, although PFGE variations within the epidemic lineages sometimes masked their genetic relatedness. In conclusion, MLVA represents a promising cost-effective first-line tool in *B. cenocepacia* surveillance.

The *Burkholderia cepacia* complex (BCC), which includes 18 closely related species, is mainly involved in pulmonary infections in patients with cystic fibrosis (CF), although it is also occasionally recovered in nosocomial infections and/or from immunocompromised hosts. BCC infection in CF patients is associated with accelerated decline of lung function, cepacia syndrome, and poor posttransplantation outcome. The most frequently recovered species of the BCC are *B. cenocepacia* and *Burkholderia multivorans*, which have been found to be responsible for large outbreaks among CF patients, prompting the implementation of infection control measures and epidemiological surveillance. *B. cenocepacia* consists of four *recA* subgroups: IIIA, which includes the Edinburgh-Toronto electrophoretic type 12 (ET-12) epidemic lineage (in Canada and the United Kingdom) (1); IIIB, which includes the PHDC epidemic lineage (in the United States) (2) and the Midwest clone (3); IIIC, which is exclusively environmental; and IIID, which was reported to be responsible for 50% of the BCC infections in an Italian CF center (4).

Putative transmissibility markers have been identified within *B. cenocepacia* epidemic strains. The *cbIA* pilin gene is characteristic of ET-12 strains (5). The *B. cepacia* epidemic strain marker (BCESM), which is part of a pathogenicity island, was identified in ET-12 strains and in other epidemic lineages but also in unique clinical or environmental strains (6, 7), whereas the PHDC lineage is BCESM negative. Finally, the IS1363 insertion sequence was demonstrated to be characteristic of the PHDC and ET-12 lineages (8). Thus, some epidemic lineages are associated with genetic

markers, but none of these proved to be universal, and epidemiological surveillance still requires genotyping studies.

Several typing methods have been developed to trace BCC organisms. Among them, pulsed-field gel electrophoresis (PFGE) and multilocus sequence typing (MLST) are currently considered the gold standard methods for epidemiological typing and population genetics studies. PFGE has been extensively used to characterize outbreaks but is a rather laborious technique that requires 9

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days to generate typing results. Another limitation of PFGE typing is the difficulty in comparing the fingerprints between laboratories, especially when different protocols and similarity thresholds for clonal delineation are used. MLST offers the advantage of being a truly portable method that allows the generation of coded types and easily accessible databases. However, MLST costs are too high for routine use in the clinical microbiology laboratory or for the characterization of large numbers of BCC strains in national surveillance programs. Additional typing methods combining good interlaboratory reproducibility with high discrimination, meaningful epidemiological inference, and low cost are still needed.

Tandem repeats (TR) consist of consecutive occurrences of a more or less identical DNA repeat unit. They are found in the chromosomes of most organisms, from prokaryotes to eukaryotes, and some TR show intraspecies polymorphisms. Although the biological function and evolution mechanism(s) of these loci with variable-number tandem repeats (VNTRs) are not yet fully understood, they have found several practical applications in epidemiological typing. Multilocus VNTR analysis (MLVA) is one of the most promising techniques for tracing the epidemiology of bacteria and has become the reference typing method in several bacterial species (9–13). In a typical MLVA assay, a predefined number of VNTRs is analyzed by PCR amplification, followed by electrophoresis to estimate the size of the PCR products. A code reflecting the number of repeats at each locus can be determined for individual strains. Therefore, MLVA is a cost-effective typing method that provides portable results, with the MLVA codes consisting of an unequivocal string of numbers that can easily be exchanged between laboratories and made accessible via internet databases (12). MLVA codes can also be deduced from whole-genome sequence data (14). An MLVA scheme has been developed to type *Burkholderia pseudomallei* (15, 16).

The aim of this study was to establish an MLVA scheme for *B. cenocepacia*. For this purpose, we searched for the presence of shared VNTR markers in the complete genome sequences of four *B. cenocepacia* strains and set up a standard MLVA protocol based on simple PCR and agarose gel electrophoresis. The overall performances of the MLVA method were assessed in a study of a selected collection of *B. cenocepacia* strains and compared with those of reference typing methods, including PFGE and MLST.

## MATERIALS AND METHODS

**Bacterial strains.** A total of 109 strains were used in the present study, which included 25 reference strains and 84 strains from the French Observatoire *Burkholderia cepacia* (OBC) collection. The 25 reference strain panel included seven *B. cenocepacia* IIIA strains (J2315, LMG18827, LMG18826, LMG18863, LMG6981, LMG18828, and LMG6986), nine *B. cenocepacia* IIIB strains (AU1054, LMG16659, LMG18829, LMG18830, LMG13011, LMG14271, LMG14274, LMG14276, and LMG16654), eight *B. cenocepacia* IIIC strains (LS2.4, Aus26, Aus12, Aus31, Aus32, Aus34, Aus13, and WS11.7, referred to in the LMG collection as LMG19233, LMG19247, LMG19243, LMG19244, LMG19248, LMG19242, LMG19240, and LMG19232, respectively), and one *B. cenocepacia* IIID strain (LMG21461). Strain AU1054 (PHDC lineage) was kindly provided by John LiPuma, University of Michigan, Ann Arbor, MI, USA, and strain J2315 (ET-12 lineage) by Catherine Doherty, University of Edinburgh, Edinburgh, Great Britain; other clinical strains were obtained from the LMG collection, Ghent, Belgium. The environmental *B. cenocepacia* IIIC strains were kindly provided by Jacques Balandreau, UMR 5557 CNRS,

Université Claude Bernard, Lyon, France. Nine reference strains were part of the diagnostically and experimentally useful panel of strains of the BCC (17). The panel of 84 nonredundant clinical strains from the OBC collection included epidemiologically related and unrelated strains (Table 1; see also Table S2 in the supplemental material) and comprised 72 CF strains from 20 CF centers, as well as 12 non-CF strains. These 84 strains had been tested with the species-specific *recA*-based PCRs described by Mahenthiralingam et al. (18), and 46 were assigned to *B. cenocepacia* IIIA and 37 to *B. cenocepacia* IIIB. One strain was species-specific PCR negative and was subsequently identified as *B. cenocepacia* IIID by means of *recA* sequencing.

Also, 39 clinical isolates recovered from 14 patients (two to four successive isolates per patient sampled over a mean period of 4.7 years) were included to assess the *in vivo* stability of the MLVA results (longitudinal stability panel; see Table S1 in the supplemental material).

**Selection of VNTR markers.** The annotated genome sequences of strain AU1054 (GenBank accession no. NC\_008060, NC\_008061, and NC\_008062), J2315 (NC\_011000, NC\_011001, and NC\_011002), and strain MCO-3 (NC\_010508, NC\_010512, and NC\_010515) were inspected for the presence of potential VNTR loci by using the strain comparison tool available at the Microorganisms Tandem Repeats Database (<http://tandemrepeat.u-psud.fr/>). Ten VNTRs were selected on the basis of polymorphisms and repeat size (Table 2). The full locus names include the following allele calling convention: repeat unit size, expected PCR product size with the indicated primers in the first published complete genome sequence (J2315), and corresponding repeat unit number assigned by convention (for example, BcenM01\_49bp\_399bp\_4U). Abbreviated names (BcenM01) are used here for simplification.

**Nomenclature and description of MLVA profiles.** The expected amplicon size, repeat length, and number of repetitions were determined in reference genomes of the AU1054, MCO-3, and J2315 strains available at the National Center for Biotechnology Information (see <http://www.ncbi.nlm.nih.gov/sites/genome>), using the PCR primer BLAST tool from the Microorganisms Tandem Repeats Database and an *in silico* MLVA typing tool (see <http://mlva.u-psud.fr/>). The number of repeats was calculated according to an excess approximation in order to consider also the non-integer number of repeats in the allele counts, as explained in Vergnaud and Pourcel (10, 13). The length of the individual repeats within some of the tandem repeats showed variations (Table 2), but repeat copy numbers were unambiguously derived from size estimates, mostly due to large repeat units and moderate repeat copy numbers. The allelic profile was defined as the number of repeats at each VNTR locus included in the MLVA scheme. When no amplification was repeatedly observed at a given locus, the value “0” was assigned to the corresponding allele.

**Template DNA for PCRs.** Bacterial thermolysates were prepared from overnight cultures using a rapid procedure; four colonies were suspended in 200  $\mu$ l of sterile water, incubated 10 min at 95°C, frozen at –20°C, and thawed. The supernatant was recovered after centrifugation and stored at –20°C.

**PCR amplification of VNTR loci.** The primers used are described in Table 2. PCR was performed in a reaction volume of 25  $\mu$ l containing 1 $\times$  PCR buffer, 1.5 mM MgCl<sub>2</sub>, 1 M betaine (Sigma-Aldrich, Saint-Quentin-Fallavier, France), 1 U of Taq DNA polymerase (Eurobio, Les Ulis, France), 200  $\mu$ M each deoxynucleoside triphosphate (Eurobio), 0.2  $\mu$ M each primer (Eurofins MWG Operon, Les Ulis, France), and 2.5  $\mu$ l of DNA template. Amplification was carried out using a VWR Doppio thermal cycler, under the following conditions: initial denaturation at 96°C for 4 min, 30 cycles of denaturation at 96°C for 30 s, annealing at 60°C for 1 min, and elongation at 72°C for 1 min, and a final elongation step at 72°C for 5 min. One microliter of PCR product was separated in 2% (for BcenM01, BcenM02, BcenM03, BcenM04, BcenM07, and BcenM10), 1.5% (for BcenM05 and BcenM09), or 1% (BcenM06 and BcenM08) (wt/vol) agarose gels (Resophor; Eurobio) in Tris-borate-EDTA (TBE) buffer (Euromedex, Souffelweyersheim, France). The PCR products from

TABLE I Characteristics and genotypes of the epidemiologically related panel of strains used for the evaluation of *in vivo* transverse stability and of the epidemiological concordance of MLVA (*n* = 39)

Strain <sup>a</sup>	<i>recA</i> subgroup lineage	Source	Location	PCR ribotype	PFGE	MLVA profile <sup>c</sup>	MLVA type	MLST results <sup>d</sup>										
								<i>atpD</i>	<i>gltB</i>	<i>gyrB</i>	<i>recA</i>	<i>lepA</i>	<i>p<sub>ha</sub>C</i>	<i>trpB</i>	ST			
French outbreak-related strains ( <i>n</i> = 25)																		
OBC01_52	IIIA French type 3	CF patient	France (Bretagne)	A	BD	3, 3, 2, 3, 2, 1, 2, 9, 0, 3	A1	16	11	10	14	11	6	79	32			
OBC02_21	( <i>n</i> = 8) <sup>b</sup>	CF patient		A	BD	3, 3, 2, 3, 2, 1, 2, 9, 0, 3	A1											
OBC15_34		CF patient		A	BE	3, 3, 2, 3, 2, 1, 2, 9, 0, 3	A1	16	11	10	14	11	6	79	32			
OBC21_17		CF patient		A	BD	3, 0, 2, 3, 2, 1, 2, 9, 0, 3	A24											
OBC02_79		CF patient	France (PACA)	A	BD	3, 3, 2, 3, 2, 1, 2, 9, 0, 3	A1											
OBC03_03		CF patient		A	BD	3, 3, 2, 3, 2, 1, 2, 9, 0, 3	A1											
OBC01_72		CF patient	France (Lorraine)	A	BD	3, 3, 2, 3, 2, 1, 2, 9, 0, 3	A1											
OBC01_01		CF patient		A	BD	3, 3, 2, 3, 2, 1, 2, 9, 0, 3	A1											
OBC05_12	IIIB French type 11	CF patient	France (PACA)	D	C	2, 6, 4, 2, 3, 4, 2, 9, 4, 6	B2	16	16	266	191	68	41	13	279			
OBC05_36	( <i>n</i> = 9) <sup>b</sup>	CF patient		D	C	2, 6, 4, 2, 3, 4, 2, 9, 4, 6	B2											
OBC17_29		CF patient		D	C	2, 6, 4, 2, 3, 4, 2, 9, 4, 6	B21											
OBC01_06		CF patient	France (Alsace/Lorraine)	D	C	2, 6, 4, 2, 3, 4, 2, 9, 4, 6	B2											
OBC08_67		CF patient		D	C	2, 6, 4, 2, 3, 4, 2, 9, 4, 6	B2											
OBC07_33		CF patient		D	C	2, 6, 4, 2, 3, 4, 2, 9, 4, 6	B2											
OBC05_74		CF patient	France (Bretagne)	D	C	2, 6, 4, 2, 3, 4, 2, 9, 4, 6	B2											
OBC06_46		CF patient		D	C	2, 6, 4, 2, 3, 4, 2, 9, 4, 6	B2											
OBC06_25		CF patient		D	C	2, 6, 4, 2, 3, 4, 2, 9, 4, 6	B2											
OBC12_32	IIIB French type 2	CF patient	France (Auvergne)	L	DZ	3, 5, 4, 4, 3, 4, 3, 8, 4, 5	B1	23	134	57	15	93	8	14	122			
OBC07_30	( <i>n</i> = 8) <sup>b</sup>	CF patient		L	DX	3, 5, 4, 4, 3, 4, 3, 8, 4, 5	B1	23	134	57	15	93	8	14	122			
OBC12_36		CF patient		L	DX	3, 5, 4, 4, 3, 4, 3, 8, 4, 5	B1											
OBC12_38		CF patient		L	DX	3, 5, 4, 4, 3, 4, 3, 8, 4, 5	B1											



TABLE 2 Characteristics and performances of the 10 selected VNTR loci

VNTR locus	Size of repeat unit (bp)	Product size (bp) (no. of repeats) in strain:			Primer sequence (5' to 3') <sup>a</sup>	100 <i>B. cereus</i> strains						Total (n = 60)		
		MCO-3	J2315	AU1054		60 epidemiologically unrelated IIIA and IIIB strains (HGDI values [95% CI]) <sup>b</sup> :								
						III A (n = 49)	III B (n = 41)	III C (n = 8)	III D (n = 2)	Total (n = 100)	IIIA (n = 36)		IIIB (n = 24)	Total (n = 60)
BeenM01_49bp_399bp_4U	49	308 (2)	399 (4)	357 (3)	F: GTAGCCGAGCTCTGTAGTCTGT; R: CTACTGTGATGACCGGTGAG	0-4 (5)	0 (0)	2 (4.8)	0 (0)	0 (0)	2 (2)	0.53 (0.47-0.60)	0.59 (0.43-0.75)	0.69 (0.65-0.72)
BeenM02_15bp_207bp_3U	15	225 (4)	207 (3)	240 (5)	F: TAAGCCACGAAAGCCCGGCA; R: GCAACCGGGGATACCCGGTG	0-6 (7)	1 (2)	1 (2.4)	0 (0)	0 (0)	2 (2)	0.33 (0.15-0.50)	0.76 (0.65-0.87)	0.56 (0.42-0.70)
BeenM03_49bp_165bp_1U	49	315 (4)	165 (1)	315 (4)	F: GATGACGATCGCGGATAAT; R: AGTCCATTCGCGCGGATGA	1-8 (6)	0 (0)	0 (0)	0 (0)	0 (0)	0 (0)	0.73 (0.65-0.80)	0.66 (0.50-0.82)	0.73 (0.69-0.78)
BeenM04_49bp_229bp_3U	49	138 (1)	229 (3)	285 (4)	F: GTACTGACGGGTTTGTATCGI; R: TCATTTGGCGCGGTTTT	0-4 (5)	2 (4)	0 (0)	0 (0)	0 (0)	2 (2)	0.11 (0.00-0.24)	0.66 (0.54-0.78)	0.61 (0.47-0.73)
BeenM05_49bp_508bp_2U	49	493 (2)	508 (2)	553 (3)	F: AAATACGGCTAGTAGTCTGTG; R: TTGGACTTACGTATATGCCCGG	2-5 (4)	0 (0)	0 (0)	0 (0)	0 (0)	0 (0)	0	0.39 (0.21-0.57)	0.43 (0.33-0.52)
BeenM06_49bp_438bp_1U	49	599 (4)	438 (1)	603 (4)	F: CGCTCGACTTCGGCGGATCA; R: ACGCGAATCTGGCTCGTGA	1-4 (3)	0 (0)	0 (0)	0 (0)	0 (0)	0 (0)	0	0.54 (0.43-0.65)	0.55 (0.47-0.66)
BeenM07_17bp_140bp_2U	17	139 (2)	140 (2)	156 (3)	F: TTCAGCCGTATGGCGGGGGCG; R: TGACGGTCCGCCACCGGCA	0-4 (4)	0 (0)	1 (2.4)	8 (100)	0 (0)	9 (9)	0	0.49 (0.33-0.65)	0.42 (0.31-0.53)
BeenM08_75bp_1023bp_9U	75	1173 (11)	1023 (9)	948 (8)	F: CCGCGGTGTGTTCGGAG; R: CAGCTGTTTCGACCGGGGT	0-11 (7)	0 (0)	1 (2.4)	8 (100)	0 (0)	9 (9)	0.71 (0.66-0.76)	0.62 (0.48-0.75)	0.71 (0.66-0.77)
BeenM09_49bp_346bp_2U	49	396 (3)	346 (2)	446 (4)	F: GTGGGGGGACCAAGACGGCG; R: ATCTGGATCCGGATCTGTATGA	0-7 (6)	26 (53)	0 (0)	0 (0)	0 (0)	26 (26)	0.65 (0.56-0.75)	0.62 (0.49-0.74)	0.72 (0.69-0.76)
BeenM10_54bp_265bp_3U	54	307 (4)	265 (3)	340 (5)	F: TGGCGGGGGCTGAGGCAT; R: CCGCCCGGCTGCATGACCGC	0-6 (6)	0 (0)	3 (7.3)	4 (50)	2 (100)	9 (9)	0.05 (0.00-0.16)	0.54 (0.33-0.75)	0.59 (0.48-0.70)

<sup>a</sup> F, forward; R, reverse. Annealing temperature, 60°C.

<sup>b</sup> 95% CI, 95% confidence interval. The global HGDI values (95% CI) were 0.96 (0.93 to 0.99) for IIIA, 0.97 (0.94 to 1.00) for IIIB, and 0.98 (0.97 to 0.99) total.

strains J2315 and AU1054 (of a known expected size), and two DNA size markers, i.e., 50-bp DNA ladder and pUC mix marker 8 (Fermentas, Villebon-sur-Yvette, France), were included in each run to allow size assignment.

**Performance criteria for evaluation of MLVA typing.** MLVA was evaluated according to the Guidelines for the validation and application of typing methods for use in bacterial epidemiology (19).

The interlaboratory reproducibility of MLVA was tested on 11 of the 25 reference strains, composed of seven clinical strains (LMG16656, LMG18827, LMG18826, LMG24506, LMG6981, LMG18828, and LMG16659) and four environmental strains (LMG19232, LMG19233, LMG19243, and LMG19248). MLVA was performed at the University Paris Sud, Orsay laboratory, and in the Observatoire *B. cepacia* laboratory, Toulouse, France, and the results were compared.

Typeability (T) was assessed for each of the 10 VNTR loci tested (Table 2). Strains for which two or more VNTRs were not amplified were further submitted to *recA* sequencing in order to ascertain that their initial identification as *B. cenocepacia* by IIIA- or IIIB-specific PCR positivity was correct.

*In vivo* longitudinal stability (LS) was assessed by testing the longitudinal stability panel (see Table S1 in the supplemental material). *In vivo* transversal stability (TS) was assessed by comparing the MLVA profiles of 25 outbreak-related strains belonging to the three genotypes involved in major outbreaks in France and referred to as genotypes 2, 3, and 11 in a previous study (20) (Table 1).

Epidemiological concordance (E) was assessed by testing 39 strains, which included the abovementioned 25 outbreak-related clinical strains, six epidemiologically related strains involved in cross-transmission, four strains belonging to the ET-12 lineage, and four environmental reference strains of *B. cenocepacia* IIIC belonging to the same sequence type (ST-118).

Discriminatory power was evaluated using a panel of 60 epidemiologically unrelated clinical strains belonging to the IIIA (36 strains) and IIIB (24 strains) subgroups, which included 10 reference strains and one strain of each epidemic clone (see Table S2 in the supplemental material). The Hunter-Gaston discriminatory index (HGDI) and 95% confidence interval were calculated using the BioNumerics software version 7.1 (Applied Maths, Sint-Martens-Latem, Belgium).

The MLVA typing results were compared to the results obtained by PCR ribotyping (21) routinely used for epidemiological screening in the Observatoire *B. cepacia* laboratory and PFGE, performed as previously described (20).

Finally, the unweighted-pair group method using average linkages (UPGMA) was used for clustering analysis and dendrogram generation (BioNumerics).

***recA* sequencing and MLST.** *recA* sequencing was performed on nontypeable strains according to Mahenthiralingam et al. (18), and the sequence data were analyzed using the Basic Local Alignment Search Tool (BLAST) ([www.ncbi.nlm.nih.gov/BLAST/](http://www.ncbi.nlm.nih.gov/BLAST/)).

A selection of strains was subjected to MLST using the scheme proposed by Baldwin et al. (22). Amplification was performed in a reaction volume of 50  $\mu$ l containing 1 $\times$  PCR buffer, 1.5 mM MgCl<sub>2</sub>, 1 M betaine, 1.5 U of Taq DNA polymerase, 200  $\mu$ M each deoxynucleoside triphosphate, 0.4  $\mu$ M each primer, and 4  $\mu$ l of DNA template. Amplified DNA was purified using the QIAquick PCR purification kit (Qiagen, Courtaboeuf, France). DNA concentration and purity were assessed using spectrophotometric analysis (BioPhotometer; Eppendorf, Le Pecq, France) and DNA concentrations adjusted to 10 ng/ $\mu$ l. The sequencing reactions were performed by the genomic facility GeT-Purpan of the Genomic and Transcriptomic Platform, Génopole Toulouse-Midi-Pyrénées, using the ABI Prism BigDye Terminator version 3.1 cycle sequencing kit (Applied Biosystems, Foster City, CA) and the 3130XL genetic analyzer (Applied Biosystems). The *B. cepacia* complex MLST website (<http://pubmlst.org/bcc/>) developed by Keith Jolley and located at the University of Oxford, Oxford, United Kingdom (23) was used for the MLST analysis.

**IS1363-specific PCR.** The primers P1 (5'-GCTTAATAGGATGGTCA G-3') and P2 (5'-TCCATGACCACCGTACAACCTC-3') were those described by Liu et al. (8). PCR was performed in a reaction volume of 25  $\mu$ l containing 1 $\times$  PCR buffer, 1.5 mM MgCl<sub>2</sub>, 1 U of Taq DNA polymerase, 200  $\mu$ M each deoxynucleoside triphosphate, 0.6  $\mu$ M each primer, and 4  $\mu$ l of DNA template. DNA from strain AU1054 was used as a positive control. Amplification was carried out under the following conditions: initial denaturation for 5 min at 95°C, 30 cycles of 45 s at 95°C, 45 s at 55°C, and 45 s at 72°C, with a final extension step of 10 min at 72°C. The PCR products were separated in 1.5% (wt/vol) agarose gels (Eurobio) in TBE buffer.

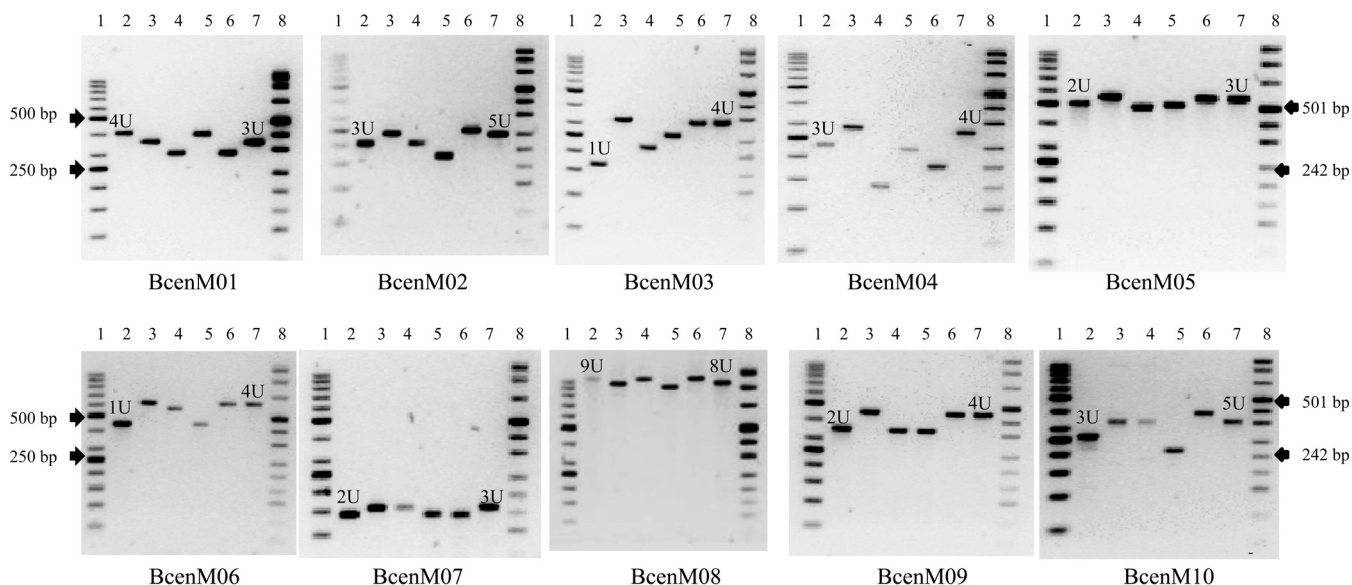
## RESULTS

**VNTR identification and interlaboratory reproducibility.** Fourteen VNTRs were identified by a genome comparison of three available sequenced strains. Ten were subsequently retained on the basis of a minimum repeat size of 15 bp and more than one allele within the 11 reference strains constituting the test panel for reproducibility (Table 2). The MLVA results were identical in both laboratories for the 11 strains.

**Typeability.** Amplification failures were observed at two, three, or four VNTRs in 18 of the 109 strains tested: 8/18 were environmental *B. cenocepacia* IIIC strains, which were all negative for at least BcenM07 and BcenM08; five were IIIA *recA*-based specific PCR positive, and subsequent *recA* sequencing identified four of them as *B. contaminans* (with at least BcenM07, BcenM08, and BcenM10 being negative) and the last one as *Burkholderia cenocepacia* IIIA; last, five were IIIB *recA*-based specific PCR positive, and subsequent *recA* sequencing identified them as belonging to the BCC6 group described by Dalmastrri et al. (24). The nine BCC6 group and *B. contaminans* strains were excluded from further analysis, which was consequently based on 100 *B. cenocepacia* strains (Table 2). Individual marker typeability was found to be  $\geq 0.98$  for six markers (BcenM01 to BcenM06) and 0.91 for three markers (BcenM07, BcenM08, and BcenM10). Last, BcenM09 amplification failure was observed in 26 of the 49 *B. cenocepacia* IIIA strains (53%), in contrast with the full BcenM09 typeability of other subgroups. The global T values for the MLVA-10 scheme were 0.43 for *B. cenocepacia* IIIA strains and 0.80 for *B. cenocepacia* IIIB strains. The low global typeability of *B. cenocepacia* IIIA strains is partly due to the overrepresentation (11/49) of ST-32 strains, for which BcenM09 amplification was never obtained. In addition, BcenM09 was not amplified in three strains sharing the PCR ribotype CE and the PFGE type DE (ST-201). Thus, considering the absence of BcenM09 amplification as a signature for ST-32 and ST-201 strains, it is possible to evaluate BcenM09 typeability as 0.76 and the global typeability of *B. cenocepacia* IIIA strains as 0.69.

**Allelic variation.** The number of alleles of the different markers ranged from three to seven within the 100 tested strains (Table 2). Of note is that all the strains of *B. cenocepacia* IIIA tested in this study ( $n = 49$ ) were monomorphic with regard to the number of repeat units of BcenM04 (three repeat units, with the exception of two amplification failures), BcenM05 (two repeat units), BcenM06 (one repeat unit), and BcenM07 (two repeat units), and they were almost monomorphic at BcenM10 (mostly three repeat units except for one strain). Figure 1 shows an example of an electrophoretic analysis of VNTR polymorphisms.

**Longitudinal stability.** The longitudinal stability panel (see Table S1 in the supplemental material) included 39 isolates from 14 patients. Nine patients were colonized with *B. cenocepacia* IIIA



**FIG 1** Electrophoretic analysis of the number of tandem repeats at each VNTR. Lane 1, 50-bp DNA ladder; lane 2, strain J2315 (LMG16656); lanes 3 to 6, four of the strains tested in the present study; lane 7, strain AU1054 (LMG24506); lane 8, pUC mix marker 8. The numbers of tandem repeats (U) are indicated for J2315 and AU1054 strains, used as controls. The arrows indicate fragment sizes of the DNA ladders.

and five with *B. cenocepacia* IIIB. The mean time between the first and the last isolate was 4.7 years (range, 1.1 to 10.2). In 12 of the 14 patients (86%), the MLVA profiles were identical over time. In two patients, a difference at BcenM08 was observed in one of the isolates tested, i.e., one amplification failure and a deletion of one repeat. Thus, an LS value of 1.00 was achieved for all markers except BcenM08 (LS, 0.90).

**Transversal stability and epidemiological concordance.** Twenty-five French outbreak-related strains, involving three epidemic lineages, were tested. All strains within each epidemic lineage harbored the same MLVA profile, with the exception of two amplification failures (at BcenM02 and BcenM10). Thus, a TS value of 1.00 was achieved for all markers, except BcenM02 and BcenM10 (TS, 0.96).

Two pairs of French epidemiologically related strains showed the same MLVA type, and within a third pair, the only difference was that BcenM04 failed to be amplified in one isolate. Similarly, four strains belonging to the *B. cenocepacia* IIIA ET-12 lineage and four ST-118 environmental *B. cenocepacia* IIIC strains were identical. In conclusion, epidemiological concordance assessed using 39 epidemiologically related strains showed an E value of 0.92 (Table 1).

**Discriminatory power.** The discriminatory power of the MLVA scheme was assessed using a panel of 60 epidemiologically unrelated strains (see Table S2 in the supplemental material). The HGDI values and 95% confidence intervals are reported in Table 2. An HGDI value of 0.98 was observed when the 10 VNTRs were combined. Five of the 10 markers (BcenM01, BcenM03, BcenM04, BcenM08, and BcenM09) were highly discriminating (HGDI, >0.6), whereas the remaining markers provided HGDI values of 0.42 to 0.59. A marked difference was observed between the IIIA and IIIB strains. Within the IIIA strains, only three markers (BcenM03, BcenM08, and BcenM09) provided HGDI values of >0.6, whereas five markers provided HGDI values of <0.3. However, the combined index was similar for both groups, reflect-

ing the marker complementarities. The numbers of IIIC and IIID strains included in the present study were low (eight and two strains, respectively), and allelic variation was observed with only two markers (BcenM02 and BcenM05 and BcenM05 and BcenM08, respectively).

**Comparison of MLVA and PFGE within *B. cenocepacia* IIIA and IIIB strains.** PFGE results were available for 82 strains, which included 44 *B. cenocepacia* IIIA and 38 *B. cenocepacia* IIIB strains, and these were compared with the results of MLVA typing. Strains exhibiting contradictory PFGE and MLVA classifications were submitted to MLST. The 82 strains were distributed into 41 MLVA types and 47 PFGE types.

Fifty-three strains were classified into 12 shared MLVA types that included 2 to 10 strains, as well as 24 PFGE types. Of the 12 shared MLVA types, two (A5 and B2) included epidemiologically related strains only, three (A1, A3, and B1) included epidemiologically related and unrelated strains, six (A4, A12, A20, A18, B5, and B8) included epidemiologically unrelated strains only, and the last one (B4), had strains LMG14274, LMG14276, and LMG13011, which were recovered from CF patients in Belgium. The PFGE results were in agreement with the MLVA classifications within six of these 12 shared MLVA types, i.e., A12, A20, A5, B2, B4, and B5, which encompassed 22 strains. Conversely, several PFGE types were differentiated within the six remaining shared MLVA types consisting of 31 strains. The 25 strains belonging to MLVA types A1, B1, and A3 were distributed into three, five, and four PFGE types, respectively, whereas MLST analysis grouped the MLVA type A1 strains into ST-32, MLVA type B1 strains into ST-122, and MLVA type A3 strains into three very close STs (ST-276, ST-234, and ST-869, with two nucleotides difference). Two pairs of *B. cenocepacia* IIIA epidemiologically unrelated strains shared a PCR ribotype (FK and CU, respectively) and MLVA types (A4 and A18, respectively) but were classified into different PFGE types and different STs, mainly due to differences in the *gyrB* allele. Finally, two strains of *B. cenocepacia* IIIB classified into the B8



MLVA type were different by all the other techniques. Within the nine shared MLVA types that included epidemiologically unrelated strains, four match globally distributed STs (MLVA type A1/ST-32, MLVA type A3/ST-234, MLVA type A20/ST-241, and MLVA type B1/ST-122), and two match STs reported in another country (MLVA type A12/ST-201 and MLVA type A18/ST-665 in Italy and New Zealand, respectively) (<http://pubmlst.org/bcc>).

Twenty-nine strains were classified into 29 unique MLVA types and 24 PFGE types. Twenty of these 29 strains were classified into unique MLVA types and unique PFGE types. Three strains with unique MLVA types (21\_17/MLVA type A24, 17\_29/MLVA type B21, and 22\_78/MLVA type A26) classified into shared PFGE types (BD, C, and HB, respectively) were part of the epidemiologically related panel and differed from the other strains harboring the same PFGE type by a "0" allele at one locus. Two pairs of epidemiologically unrelated strains (12\_03 and 22\_59, and 05\_12 and 9\_54) were classified into shared PFGE types (GO and C, respectively) and shared STs (844 and 279, respectively). One pair of epidemiologically unrelated strains (20\_77 and 20\_78) was classified into the same PFGE type (FY) and two different STs (ST-760 and ST-845, 8 nucleotides difference).

In conclusion, MLVA classification was in agreement with PFGE and/or MLST in 69 of 82 strains (84%).

**Population analysis.** UPGMA analysis showed that MLVA classification was in agreement with *recA* classification, with the exception of two IIIB strains. A cutoff of 85% was used to delineate clusters, corresponding to a maximum of one difference among the 10 loci (Fig. 2).

Four clusters designated IIIB-C1 to -C4 were delineated within *B. cenocepacia* IIIB strains, which included 26 of the 41 tested strains. MLVA clusterization within the *B. cenocepacia* IIIB isolates fit with epidemiological and/or genetic relatedness: cluster IIIB-C1 consisted of Belgian CF strains, cluster IIIB-C2 of ST-279 strains (French epidemic type 11, reported only in France to date), cluster IIIB-C3 of ST-122 strains, including the PHDC North American epidemic strain and outbreak-related CF strains (French epidemic type 2), and cluster IIIB-C4 of epidemiologically unrelated strains (two CF and one non-CF), which exhibited the same PFGE type. Following the identification of French strains harboring the same MLVA type as strain AU1054/LMG24506 (PHDC *B. cenocepacia* IIIB lineage), characterized by the presence of IS1363, an IS1363-specific PCR was performed in *B. cenocepacia* IIIB-PCR-positive strains. Strain OBC08\_30, which was classified in cluster IIIB-C3 but belongs to ST-37 (10 nucleotide and 3 allele differences with ST-122) was IS1363 positive. Of note is that five *B. cenocepacia* IIIB strains were found to be IS1363 positive outside the IIIB-C3 cluster.

Six clusters designated C1 to C6 were delineated within the *B. cenocepacia* IIIA strains, which included 35 of the 49 strains tested. MLVA clusterization within the *B. cenocepacia* IIIA isolates fit with epidemiological and/or genetic relatedness for five of the six clusters: cluster IIIA-C1 was composed of ET-12 strains (and of a French CF strain), cluster IIIA-C2 and IIIA-C5 of groups of strains belonging to very close STs (one to two nucleotides difference), IIIA-C4 of strains classified into the same PCR ribotype and PFGE type, and IIIA-C6 of ST-32 strains (French epidemic type 3). On the contrary, cluster IIIA-C3 was heterogeneous (six strains, four PCR ribotypes, five PFGE types, and five markedly different STs).

Lowering the cutoff to 58% classified nearly all the *B. cenoce-*

*pacia* IIIA strains into the same group but delineated five groups within the *B. cenocepacia* IIIB strains.

## DISCUSSION

BCC organisms are recognized as important pathogens in CF. Due to their ability to spread from patient to patient, they have been involved in large outbreaks. While PCR-based methods and PFGE have successfully been applied to local epidemiology, large-scale analyses require more portable methods, such as MLST or MLVA. An MLST scheme based on seven housekeeping genes has been developed and applied to global epidemiological analyses (22, 25). Also, the *recA* gene included in the MLST scheme allows the discrimination of species within the BCC. MLVA is also a transferable typing method, which is increasingly used to trace bacterial epidemiology. In the present study, we established an MLVA-10 scheme for the typing of *B. cenocepacia* strains, which together with *B. multivorans* predominates in CF and is associated with highly transmissible lineages, such as the so-called ET-12, ST-32 (*B. cenocepacia* IIIA), Midwest, and PHDC (*B. cenocepacia* IIIB) lineages. Within the selected markers, the size of the repeat unit varied from 15 to 75 bp, which allows for allele assignment by means of simple agarose gel electrophoresis. The performance of MLVA-10 was evaluated on a large representative collection of epidemiologically related and unrelated national and international strains.

The selected VNTRs appear to be specific for *B. cenocepacia*, as *B. contaminans* and the BCC6 group strains failed to be amplified at two to four loci when using the proposed PCR primer pairs. An examination of the sequenced genomes confirmed that the VNTRs cannot be amplified *in silico* in other members of the BCC. Moreover, these loci are not equally spread within the subgroups of the *B. cenocepacia* species. BcenM09 failed to amplify in more than half of the *B. cenocepacia* IIIA strains tested, whereas it was amplified in all the *B. cenocepacia* IIIB, IIIC, and IIID strains tested. BcenM07 and BcenM08 failed to amplify in the eight *B. cenocepacia* IIIC strains tested, whereas they were amplified in all but one of the 92 strains belonging to other subgroups. Nevertheless, the absence of BcenM09 may be considered a marker for some *B. cenocepacia* lineages (ST-32 and ST-201), and the fact that *B. cenocepacia* IIIC strains are not fully typeable with the proposed scheme is not expected to impact the performance of MLVA for the typing of clinical strains, since subgroup IIIC has until now been recovered from the natural environment only.

Stability is of the utmost importance in BCC epidemiology, since most colonizations are chronic and since patient-to-patient spread may be observed over years. We checked the stabilities of the selected VNTR loci in 14 patients by testing up to four successive isolates per patient. The MLVA patterns were stable over time in 13 patients (92.8%), and the variation observed in one patient concerned a single VNTR (BcenM08, with a loss of one repeat unit). Comparatively, the longitudinal stability of the MLST results for 41 BCC isolates from 20 patients showed no change in 15 patients (75%), strain replacement in one patient, and changes due to recombination events in four patients (20%) colonized with *B. multivorans* (one patient), *B. vietnamiensis* (two patients), or *B. cenocepacia* IIIA (one patient) (26). If this trend was confirmed by larger investigations, then one might speculate that allelic differences at VNTR loci in the BCC may prevent recombination, and it would explain the stability of MLVA compared to that of the MLST profiles. In order to validate the use of MLVA for

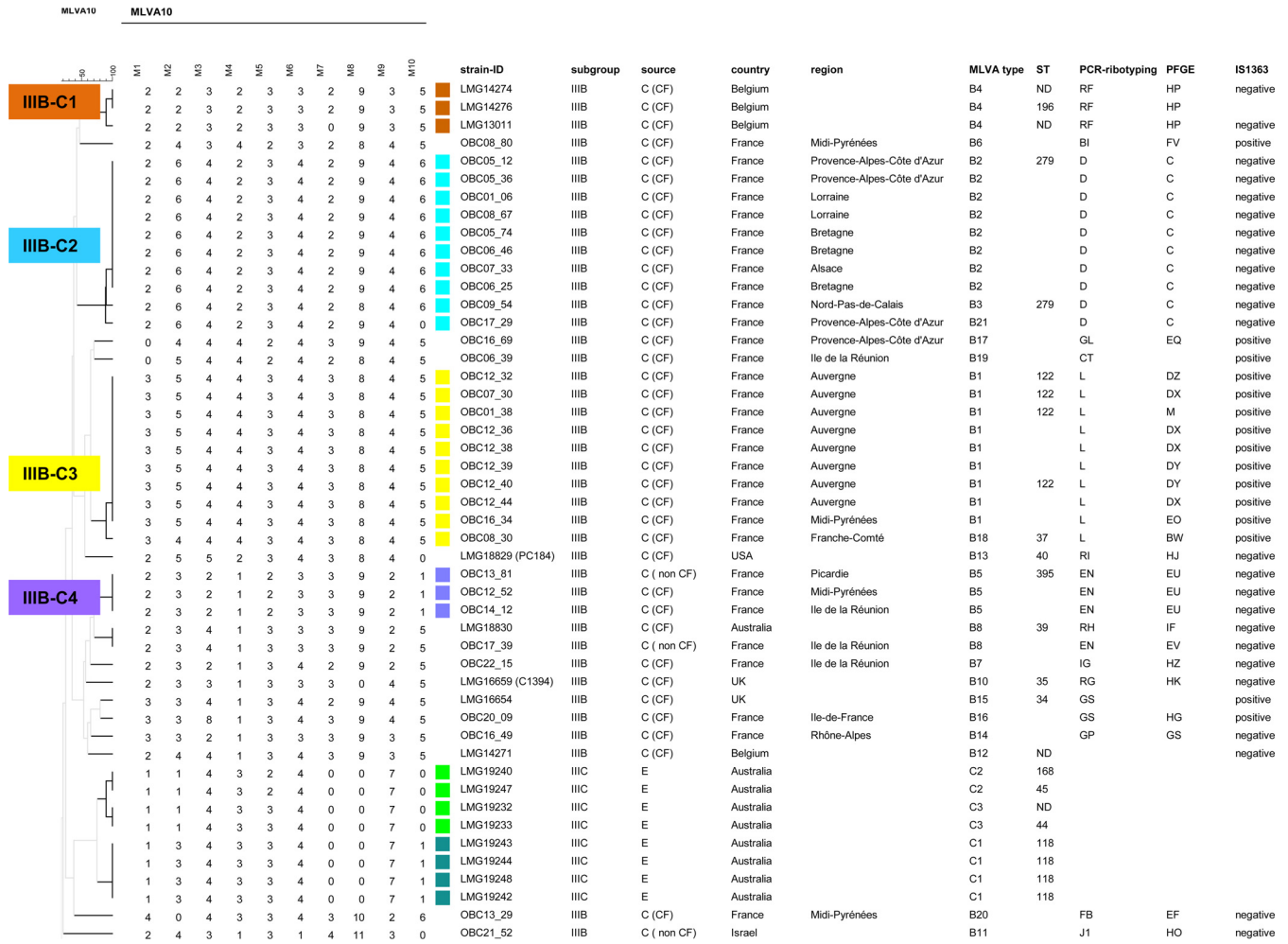


FIG 2 MLVA-based UPGMA clustering analysis of the *B. cenocepacia* strains included in the present study (cutoff, 80%). Strains classified in the same cluster are indicated using colored squares. Under the source column, there are clinical (C) and environmental (E) strains. ID, identification; CF, cystic fibrosis.

identifying outbreaks, we tested multiple strains of the three epidemic lineages of *B. cenocepacia*, i.e., ST-32, ST-279, and ST-122, responsible for outbreaks in one (ST-122) or several (ST-32 and ST-279) French CF centers in the late 1980s to the early 1990s. We included strains from each of the CF centers, collected over periods of 11 to 22 years, and demonstrated the long-term transversal stability of the MLVA profiles, with the exception of two amplification failures. Of note is that a loss of one BcenM08 repeat unit was observed in one outbreak-unrelated ST-279 strain (OBC09\_54). Thus, the marker BcenM08 might be slightly less stable than the other markers, since it also was the single marker to vary in one patient who was included in the longitudinal study. Whereas the PFGE types were identical within all the ST-279 strains tested, two, two, and five PFGE types were observed within the globally distributed lineage ST-32, ST 234, and ST-122 (PHDC) strains, respectively. PFGE pattern variability was previously reported by Drevinek and Mahenthiralingam (27) within the ST-32 Czech strains, and it was attributed to IS-mediated genomic rearrangements. Similar variability was reported by Campana et al. (28) within the Italian PHDC strains. It can be hypothesized that due to the presence of several copies of IS1363 in PHDC strains (8), IS movements might also interfere with mac-

rorestriction-based types within this lineage. Interestingly, we demonstrated the presence of IS1363 in 16 of the 41 *B. cenocepacia* IIIB strains tested, six of which were different from the PHDC lineage. Lastly, the four tested ET-12 strains exhibited the same MLVA type, whereas they belong to four different STs identified within Canadian ET-12 strains. In contrast, all reported ET-12 strains in the United Kingdom were found to belong to ST-28 (29). Thus, the evolution rates and mechanisms within each major clonal complex seem different, with variable consequences according to the typing method, leading to difficulties in establishing a unique gold standard for genotyping. This is in agreement with a previous study using multilocus restriction typing (MLRT) (30), showing differences in population structures among the ET-12, PHDC, and Midwest lineages, which were attributed to various degrees of genetic recombination. Finally, the MLVA-10 signatures of other epidemic strains, i.e., LMG18829 (ST-40, Midwest clone) and LMG16659 (ST-35, Manchester epidemic clone), were shown to be specific within the collection tested. On the contrary, LMG18830 (ST-36, Sydney epidemic clone) and a non-CF strain from La Réunion Island were classified in the same MLVA type but into different PCR ribotypes, PFGE types, and MLST profiles. This might be attributed to homoplasmy, i.e., evo-

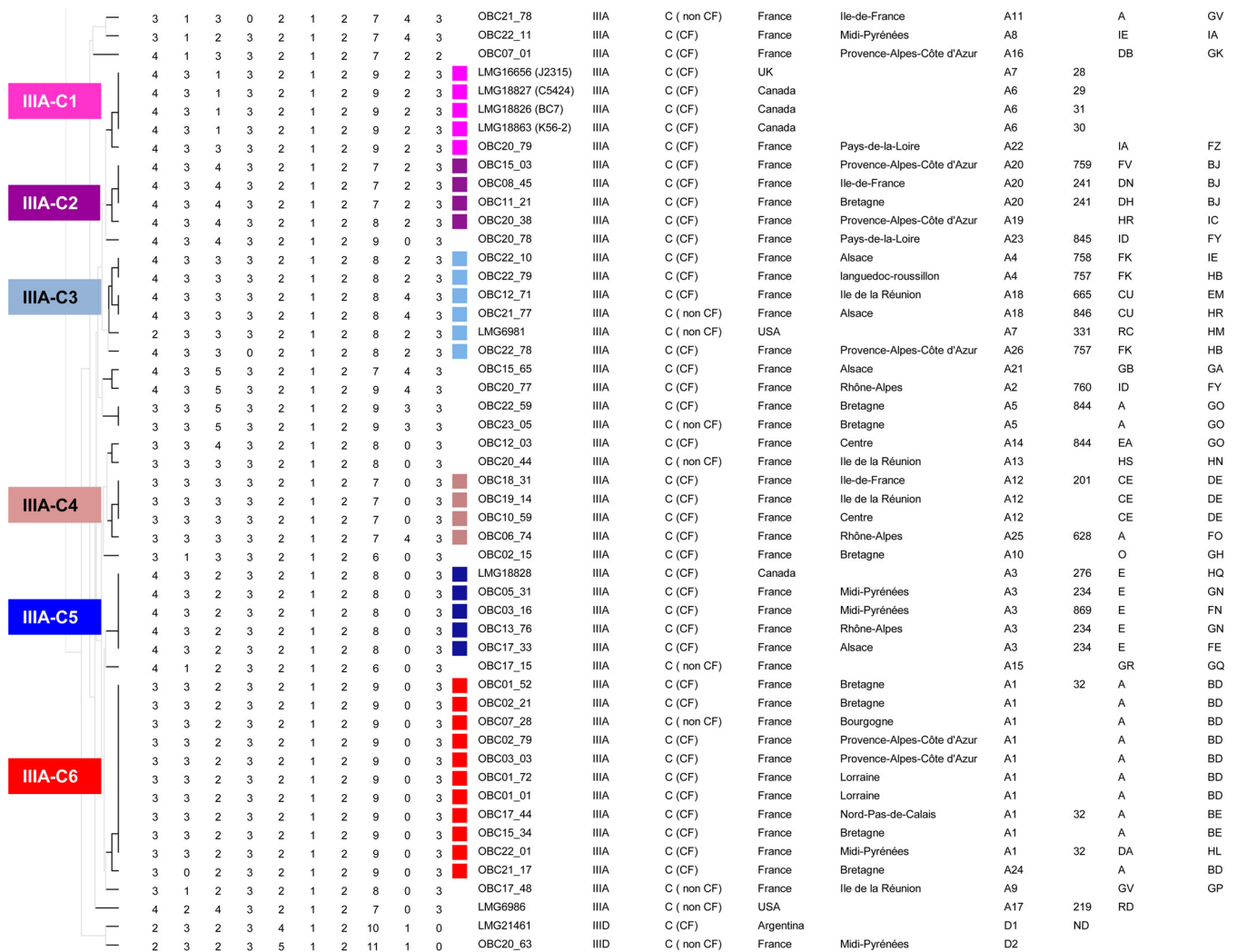


FIG 2 continued

lutionary convergence or reversion to an ancestral profile. Thus, in spite of one discordance, MLVA-10 seems a valuable tool for recognizing outbreaks and identifying widely distributed and/or epidemic lineages.

Within the 10 markers used, the number of alleles ranged from three to seven, and the diversity index (HGDI) for individual loci ranged from 0.42 to 0.73. The HGDI value for the MLVA-10 scheme was of 0.98, thus fulfilling the European Society of Clinical Microbiology and Infectious Diseases Study Group on Epidemiological Markers (ESGEM) criteria ( $>0.95$ ) (19). Compared to PFGE within epidemiologically unrelated strains, the discriminatory power of MLVA-10 seems lower than that of PFGE, which was confirmed by MLST, although the reverse situation has also been observed. Concordance of MLVA with PFGE and/or MLST results was shown to be 84% within the collection tested.

MLVA-based population structure was in agreement with *recA* classification, with the exception of two IIIB strains, which might represent a specific subgroup. Heterogeneity was higher within the *B. cenocepacia* IIIB strains than within the *B. cenocepacia* IIIA strains, since three of the 10 loci were found to be homogeneous within the *B. cenocepacia* IIIA strains, which were nearly all classi-

fied in a unique group using a cluster delineation cutoff of 58%. This may be due to a lower discriminatory power of the scheme used within IIIA strains, but it is in agreement with an MLST-based population structure. Using the matches of five out of seven loci, 198 of the 212 *B. cenocepacia* IIIA strains included in the MLST database are brought together in a large clonal complex, with ST-234 as a potential ancestral type. In contrast, the 155 *B. cenocepacia* IIIB strains are distributed into 10 clonal complexes and 81 singletons (<http://pubmlst.org/bcc/>).

In conclusion, due to the high genomic plasticity of *B. cepacia* complex organisms, recombination events or IS movements may interfere with genotyping methods and mask epidemiological relatedness. The MLVA-10 scheme developed in the present study was shown to be an appropriate tool to detect epidemic strains of *B. cenocepacia*, due to the high stability of MLVA profiles, the satisfactory discriminatory power, and the portability of the method, allowing the establishment of a Web-based database (<http://mlva.u-psud.fr>) (14). We suggest the use of MLVA as a first-line typing method for *B. cenocepacia* and further analyses of the representative MLVA profiles by means of MLST. This approach would be cost-effective, since in our laboratory, the re-

agents and sequencing costs for the characterization of one strain by means of *recA* sequence for species identification plus MLVA-10 were 36 euros compared to 112 euros for MLST analysis, with comparable hands-on time. Besides, the MLVA-10 scheme could easily be automated using fluorescent primers and a single multiplex PCR (31). Capillary electrophoresis would allow automatic size measurement of the PCR products and thus increase the accuracy of repeat number determination, as described for other bacterial species (31–33). Furthermore, some potentially informative VNTR markers that were ruled out from the present scheme due to short repeats (<15 bp) could be included to constitute a second panel with high discriminatory power, as has been proposed for other bacteria (34). Nevertheless, in contrast with MLST, which has the advantage of being usable for the whole BCC, species-specific MLVA schemes have to be developed, and the establishment of a *B. multivorans* MLVA scheme is in progress.

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## REFERENCES

- Govan JR, Brown PH, Maddison J, Doherty CJ, Nelson JW, Dodd M, Greening AP, Webb AK. 1993. Evidence for transmission of *Pseudomonas cepacia* by social contact in cystic fibrosis. *Lancet* 342:15–19. [http://dx.doi.org/10.1016/0140-6736\(93\)91881-L](http://dx.doi.org/10.1016/0140-6736(93)91881-L).
- Chen JS, Witzmann KA, Spilker T, Fink RJ, LiPuma JJ. 2001. Endemicity and inter-city spread of *Burkholderia cepacia* genomovar III in cystic fibrosis. *J Pediatr* 139:643–649. <http://dx.doi.org/10.1067/mpd.2001.118430>.
- LiPuma JJ, Mortensen JE, Dasen SE, Edlind TD, Schidlow DV, Burns JL, Stull TL. 1988. Ribotype analysis of *Pseudomonas cepacia* from cystic fibrosis treatment centers. *J Pediatr* 113:859–862. [http://dx.doi.org/10.1016/S0022-3476\(88\)80018-0](http://dx.doi.org/10.1016/S0022-3476(88)80018-0).
- Manno G, Dalmastrì C, Tabacchioni S, Vandamme P, Lorini R, Minicucci L, Romano L, Giannattasio A, Chiarini L, Bevivino A. 2004. Epidemiology and clinical course of *Burkholderia cepacia* complex infections, particularly those caused by different *Burkholderia cenocepacia* strains, among patients attending an Italian Cystic Fibrosis Center. *J Clin Microbiol* 42:1491–1497. <http://dx.doi.org/10.1128/JCM.42.4.1491-1497.2004>.
- Sun L, Jiang RZ, Steinbach S, Holmes A, Campanelli C, Forstner J, Tan Y, Riley M, Goldstein R. 1995. The emergence of a highly transmissible lineage of *cbl+* *Pseudomonas (Burkholderia) cepacia* causing CF centre epidemics in North America and Britain. *Nat Med* 1:661–666. <http://dx.doi.org/10.1038/nm0795-661>.
- Mahenthalingam E, Simpson DA, Speert DP. 1997. Identification of a novel DNA marker associated with epidemic *Burkholderia cepacia* strains recovered from patients with cystic fibrosis. *J Clin Microbiol* 35:808–816.
- Baldwin A, Sokol P, Parkhill J, Mahenthalingam E. 2004. The *Burkholderia cepacia* epidemic strain marker is part of a novel genomic island encoding both virulence and metabolism-associated genes in *Burkholderia cenocepacia*. *Infect Immun* 72:1537–1547. <http://dx.doi.org/10.1128/IAI.72.3.1537-1547.2004>.
- Liu L, Spilker T, Coenye T, LiPuma JJ. 2003. Identification by subtractive hybridization of a novel insertion element specific for two widespread *Burkholderia cepacia* genomovar III strains. *J Clin Microbiol* 41:2471–2476. <http://dx.doi.org/10.1128/JCM.41.6.2471-2476.2003>.
- Lindstedt BA. 2005. Multiple-locus variable number tandem repeats analysis for genetic fingerprinting of pathogenic bacteria. *Electrophoresis* 26:2567–2582. <http://dx.doi.org/10.1002/elps.200500096>.
- Vergnaud G, Pourcel C. 2006. Multiple locus VNTR (variable number of tandem repeat) analysis, p 83–104. In Stackebrandt E (ed), *Molecular identification, systematics, and population structure of prokaryotes*. Springer-Verlag, Berlin, Germany.
- van Belkum A. 2007. Tracing isolates of bacterial species by multilocus variable number of tandem repeat analysis (MLVA). *FEMS Immunol Med Microbiol* 49:22–27. <http://dx.doi.org/10.1111/j.1574-695X.2006.00173.x>.
- Grissa I, Bouchon P, Pourcel C, Vergnaud G. 2008. On-line resources for bacterial micro-evolution studies using MLVA or CRISPR typing. *Biochimie* 90:660–668. <http://dx.doi.org/10.1016/j.biochi.2007.07.014>.
- Vergnaud G, Pourcel C. 2009. Multiple locus variable number of tandem repeats analysis. *Methods Mol Biol* 551:141–158. [http://dx.doi.org/10.1007/978-1-60327-999-4\\_12](http://dx.doi.org/10.1007/978-1-60327-999-4_12).
- Thierry S, Tourterel C, Le Flèche P, Derzelle S, Dekhil N, Mendy C, Colaneri C, Vergnaud G, Madani N. 2014. Genotyping of French *Bacillus anthracis* strains based on 31-loci multi locus VNTR analysis: epidemiology, marker evaluation, and update of the Internet genotype database. *PLoS One* 9:e95131. <http://dx.doi.org/10.1371/journal.pone.0095131>.
- U'Ren JM, Hornstra H, Pearson T, Schupp JM, Leadem B, Georgia S, Sermawan RW, Keim P. 2007. Fine-scale genetic diversity among *Burkholderia pseudomallei* soil isolates in Northeast Thailand. *Appl Environ Microbiol* 73:6678–6681. <http://dx.doi.org/10.1128/AEM.00986-07>.
- Yen MWS, Lisanti O, Thibault F, San TS, Kee LG, Hilaire V, Jiali L, Neubauer H, Vergnaud G, Ramisse V. 2009. Validation of ten new polymorphic tandem repeat loci and application to the MLVA typing of *Burkholderia pseudomallei* isolates collected in Singapore from 1988 to 2004. *J Microbiol Methods* 77:297–301. <http://dx.doi.org/10.1016/j.mimet.2009.03.005>.
- Mahenthalingam E, Coenye T, Chung JW, Speert DP, Govan JR, Taylor P, Vandamme P. 2000. Diagnostically and experimentally useful panel of strains from the *Burkholderia cepacia* complex. *J Clin Microbiol* 38:910–913.
- Mahenthalingam E, Bischof J, Byrnes S, Radomski C, Davies J, Av-Gay Y, Vandamme P. 2000. DNA-based diagnostic approaches for identification of *Burkholderia cepacia* complex, *Burkholderia vietnamiensis*, *Burkholderia multivorans*, *Burkholderia stabilis*, and *Burkholderia cepacia* genomovars I and III. *J Clin Microbiol* 38:3165–3173.
- van Belkum A, Tassios P, Dijkshoorn L, Haeggman S, Cookson B, Fry NK, Fussing V, Green J, Feil E, Gerner-Smidt P, Brisse S, Struelens M, European Society of Clinical Microbiology and Infectious Diseases (ESCMID) Study Group on Epidemiological Markers (ESGEM). 2007. Guidelines for the validation and application of typing methods for use in bacterial epidemiology. *Clin Microbiol Infect* 13(Suppl 3):1–46. <http://dx.doi.org/10.1111/j.1469-0691.2007.01786.x>.
- Segonds C, Bingen E, Couetdic G, Mathy S, Brahimi N, Marty N, Plesiat P, Michel-Briand Y, Chabanon G. 1997. Genotypic analysis of *Burkholderia cepacia* isolates from 13 French cystic fibrosis centers. *J Clin Microbiol* 35:2055–2060.
- Dasen SE, LiPuma JJ, Kostman JR, Stull TL. 1994. Characterization of PCR-ribotyping for *Burkholderia (Pseudomonas) cepacia*. *J Clin Microbiol* 32:2422–2424.
- Baldwin A, Mahenthalingam E, Thickett KM, Honeybourne D, Maiden MC, Govan JR, Speert DP, Lipuma JJ, Vandamme P, Dowson CG. 2005. Multilocus sequence typing scheme that provides both species and strain differentiation for the *Burkholderia cepacia* complex. *J Clin Microbiol* 43:4665–4673. <http://dx.doi.org/10.1128/JCM.43.9.4665-4673.2005>.
- Jolley KA, Chan MS, Maiden MC. 2004. mlstNet—distributed multilocus sequence typing (MLST) databases. *BMC Bioinformatics* 5:86. <http://dx.doi.org/10.1186/1471-2105-5-86>.
- Dalmastrì C, Baldwin A, Tabacchioni S, Bevivino A, Mahenthalingam E, Chiarini L, Dowson C. 2007. Investigating *Burkholderia cepacia* complex populations recovered from Italian maize rhizosphere by multilocus sequence typing. *Environ Microbiol* 9:1632–1639. <http://dx.doi.org/10.1111/j.1462-2920.2007.01273.x>.
- Baldwin A, Mahenthalingam E, Drevinek P, Pope C, Waine DJ, Henry DA, Speert DP, Carter P, Vandamme P, LiPuma JJ, Dowson CG. 2008. Elucidating the global epidemiology of *Burkholderia multivorans* in cystic fibrosis by multilocus sequence typing. *J Clin Microbiol* 46:290–295. <http://dx.doi.org/10.1128/JCM.01818-07>.
- Waine DJ, Henry DA, Baldwin A, Speert DP, Honeybourne D, Mahenthalingam E, Dowson CG. 2006. Reliability of multilocus sequence typing of the *Burkholderia cepacia* complex in cystic fibrosis. *J Cyst Fibros* 6:215–219. <http://dx.doi.org/10.1016/j.jcf.2006.10.003>.
- Drevinek P, Mahenthalingam E. 2010. *Burkholderia cenocepacia* in cys-

- tic fibrosis: epidemiology and molecular mechanisms of virulence. *Clin Microbiol Infect* 16:821–830. <http://dx.doi.org/10.1111/j.1469-0691.2010.03237.x>.
28. Campana S, Taccetti G, Ravenni N, Favari F, Cariani L, Sciacca A, Savoia D, Collura A, Fiscarelli E, De Intinis G, Busetti M, Cipolloni A, d'Aprile A, Provenzano E, Collebrusco I, Frontini P, Stassi G, Trancassini M, Tovagliari D, Lavitola A, Doherty CJ, Coenye T, Govan JR, Vandamme P. 2005. Transmission of *Burkholderia cepacia* complex: evidence for new epidemic clones infecting cystic fibrosis patients in Italy. *J Clin Microbiol* 43:5136–5142. <http://dx.doi.org/10.1128/JCM.43.10.5136-5142.2005>.
  29. Mahenthiralingam E, Baldwin A, Dowson CG. 2008. *Burkholderia cepacia* complex bacteria: opportunistic pathogens with important natural biology. *J Appl Microbiol* 104:1539–1551. <http://dx.doi.org/10.1111/j.1365-2672.2007.03706.x>.
  30. Coenye T, LiPuma JJ. 2002. Multilocus restriction typing: a novel tool for studying global epidemiology of *Burkholderia cepacia* complex infection in cystic fibrosis. *J Infect Dis* 185:1454–1462. <http://dx.doi.org/10.1086/340279>.
  31. Sobral D, Le Cann P, Gerard A, Jarraud S, Lebeau B, Loisy-Hamon F, Vergnaud G, Pourcel C. 2011. High-throughput typing method to identify a non-outbreak-involved *Legionella pneumophila* strain colonizing the entire water supply system in the town of Rennes, France. *Appl Environ Microbiol* 77:6899–6907. <http://dx.doi.org/10.1128/AEM.05556-11>.
  32. Sobral D, Mariani-Kurkdjian P, Bingen E, Vu-Thien H, Hormigos K, Lebeau B, Loisy-Hamon F, Munck A, Vergnaud G, Pourcel C. 2012. A new highly discriminatory multiplex capillary-based MLVA assay as a tool for the epidemiological survey of *Pseudomonas aeruginosa* in cystic fibrosis patients. *Eur J Clin Microbiol Infect Dis* 31:2247–2256. <http://dx.doi.org/10.1007/s10096-012-1562-5>.
  33. Sobral D, Schwarz S, Bergonier D, Brisabois A, Feßler A, Gilbert F, Kadlec K, Lebeau B, Loisy-Hamon F, Treilles M, Pourcel C, Vergnaud G. 2012. High throughput multiple locus variable number of tandem repeat analysis (MLVA) of *Staphylococcus aureus* from human, animal and food sources. *PLoS One* 7:e33967. <http://dx.doi.org/10.1371/journal.pone.0033967>.
  34. Scholz HC, Vergnaud G. 2013. Molecular characterisation of *Brucella* species. *Rev Sci Tech* 32:149–162.